

# Smooth modeling of covariate effects in bisulfite sequencing-derived measures of DNA methylation



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Please feel free to interrupt and ask questions at any time during the talk!

- ▶ Background and motivation
- ▶ †New method 1: SOMNiBUS (SmOoth ModeliNg of BisUlfinite Sequencing)
- ▶ ‡New method 2: dSOMNiBUS (dispersion-adjusted SmOoth ModeliNg of BisUlfinite Sequencing)
- ▶ \*New method 3: sparseSOMNiBUS (SOMNiBUS with variable selection)

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† Zhao, et.al (2020). A novel statistical method for modeling covariate effects in bisulfite sequencing derived measures of DNA methylation. *Biometrics*. Early-View

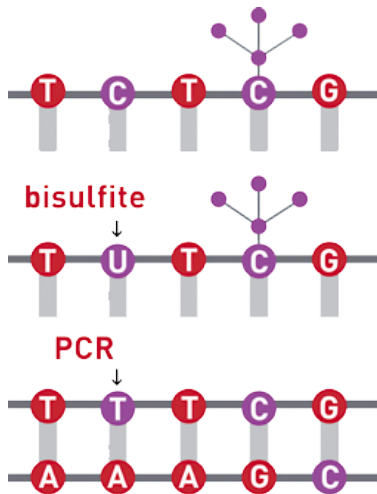
‡ Zhao, et.al (2020+). Detecting differentially methylated regions in bisulfite sequencing data using quasi-binomial mixed models with smooth covariate effect estimates.

\* Zhao, et.al (2020+). In preparation



- ▶ change gene expression without changing DNA sequence
- ▶ can be altered by age, diet, stress and environmental exposures
- ▶ Localized abnormal methylation is a characteristic feature of many diseases

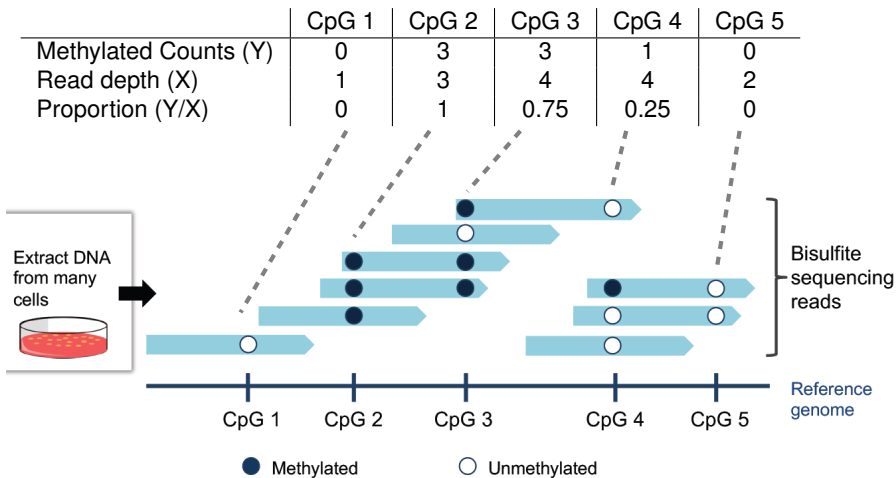
# Bisulfite Sequencing & Methylation



Methylated cytosines are not converted by bisulfite treatment

<https://www.diagenode.com/en/applications/dna-bisulfite-conversion>

# Sequencing-derived DNA methylation data



[http://kkorthauer.org/talks/korthauer\\_aisc\\_2018\\_static.pdf](http://kkorthauer.org/talks/korthauer_aisc_2018_static.pdf)

# Motivating datasets

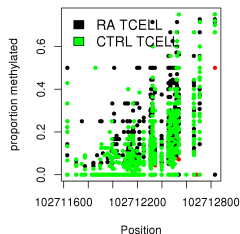
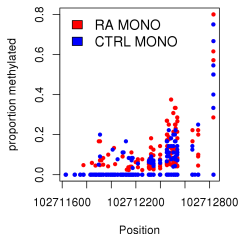
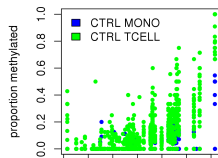
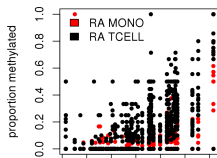
Methylation profiles of Rheumatoid Arthritis (RA) patients and controls  
(from our collaborator Dr. Marie Hudson)



- ▶ Targeted Custom Capture Bisulfite Sequencing
  - predefined genomic regions
  - 5 million CpGs
- ▶ Cell-separated blood samples

	Monocytes	T cells
RA	10	12
Controls	8	13

- ▶ Small region on chromosome 4 near *BANK1*
- ▶ 123 CpGs





## **Find associations between**

- ▶ methylation patterns in each targeted region, and
- ▶ phenotypes or covariates



- Read depth at CpGs varies substantially
  - ▶ Need a model that can use all available data
- Cell-type mixture affects observed methylation levels
  - ▶ Adjust for this in model
- Sequencing errors, e.g. bisulfite conversion error
  - ▶ Build a model allowing for error
- Local correlations in methylation levels
  - ▶ Opportunity for imputing missing data or poorly measured signals
  - ▶ Opportunity for modelling smooth effects along the genome



# Existing methods appropriate for regions



Method	regional	one-stage	count-based	read-depth variability	adjust for confounding	experimental errors
SOMNiBUS	✓	✓	✓	✓	✓	✓
BSmooth	✓			✗		
SMSC	✓			✗		✓
dmrseq	✓			✓	✓	
BiSeq	✓			✗	✓	
GlobalTest	✓	✓			✓	

**BSmooth:** Hansen, 2012

**SMSC:** Lakhali-Chaieb, 2017

**dmrseq:** Korthauer, 2018

**BiSeq:** Hebestreit, 2013

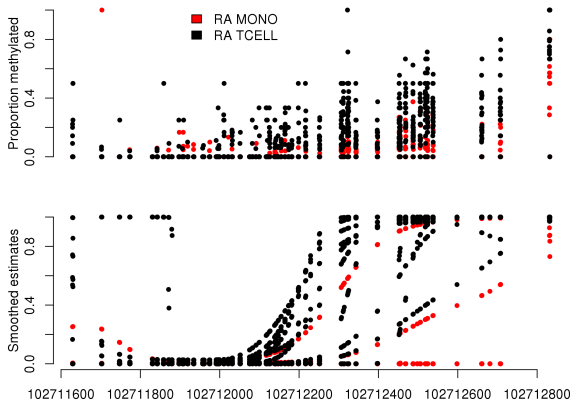
**GlobalTest:** Goeman, 2006

# An example of two-stage method

Raw data & per-sample smoothed estimates



Results from SMSC (Lakhal-Chaieb, 2017)



Genomic position (in bp)



Method	regional	one-stage	count-based	read-depth variability	adjust for confounding	experimental errors
SOMNiBUS	✓	✓	✓	✓	✓	✓
BSmooth	✓			✗		
SMSC	✓			✗		✓
dmrseq	✓			✓	✓	
Biseq	✓			✗	✓	
GlobalTest	✓	✓			✓	

**Motivation:** a novel **one-stage** method that

- ▶ collapses smoothing and testing steps into a single step
- ▶ allows for experimental errors, variable read depths and test samples with a mixture of cell types
- ▶ provides **rigorous uncertainty assessment** for differentially methylated regions



- ▶ Background and motivation
- ▶ † **New method 1: SOMNiBUS (SmOoth ModeliNg of BisUlFite Sequencing)**
- ▶ ‡ New method 2: dSOMNiBUS (dispersion-adjusted SmOoth ModeliNg of BisUlFite Sequencing)

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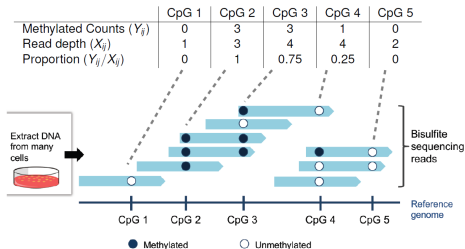
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# Notations



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- ▶  $X_{ij}$ : total number of reads aligned to CpG  $j$  from sample  $i$
- ▶  $Y_{ij}$ : **observed** methylated counts at CpG  $j$  for sample  $i$ .  $Y_{ij} = \sum_{k=1}^{X_{ij}} Y_{ijk}$
- ▶  $S_{ij}$ : **true** methylated counts at CpG  $j$  for sample  $i$ .  $S_{ij} = \sum_{k=1}^{X_{ij}} S_{ijk}$



- ▶  $t_{ij}$ : the genome position (in bp) for sample  $i$  at CpG  $j$
- ▶  $Z_{1i}, Z_{2i}, \dots, Z_{Pi}$  are the  $P$  covariates.
- ▶  $\pi_{ij}$ : the methylation proportion parameter for sample  $i$ , CpG  $j$



- ▶ Assume **known error parameters**  $\rho_0$  and  $\rho_1$ ,

$$\begin{aligned}\rho_0 &= \mathbb{P}(Y_{ijk} = 1 \mid S_{ijk} = 0) \\ \rho_1 &= \mathbb{P}(Y_{ijk} = 1 \mid S_{ijk} = 1).\end{aligned}$$

- ▶ Specify the model

$$\begin{aligned}S_{ij} \mid \mathbf{Z}_i, X_{ij} &\sim \text{Binomial}(X_{ij}, \pi_{ij}) \\ \log \left\{ \frac{\pi_{ij}}{1 - \pi_{ij}} \right\} &= \beta_0(t_{ij}) + \beta_1(t_{ij})Z_{1i} + \beta_2(t_{ij})Z_{2i} + \dots + \beta_P(t_{ij})Z_{Pi},\end{aligned}$$

- ▶ Smooth curves along the genome for
  - ▶ Overall methylation
  - ▶ Covariate effects

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<sup>†</sup>R package: <https://github.com/kaiqiong/SOMNiBUS>.



- ▶ Use splines for smoothing

$$\beta_p(t_{ij}) = \sum_{l=1}^{L_p} \alpha_{pl} B_l^{(p)}(t_{ij}) \text{ for } p = 0, 1, \dots, P.$$

- ▶ Penalize roughness of effect curves  $\beta_p(t_{ij})$ .

$$\mathcal{L}^{\text{Penalization}} = \sum_{p=0}^P \lambda_p \int (\beta_p''(t))^2 dt = \sum_{p=0}^P \lambda_p \alpha_p^T \mathbf{A}_p \alpha_p = \alpha^T \mathbf{A}_\lambda \alpha,$$

$\{\lambda_0, \lambda_1, \dots, \lambda_P\}$  are the smoothing parameters.

- ▶ Penalties go onto second derivatives
- ▶  $P + 1$  penalization parameters for  $P$  covariates



**E step:** Calculate  $\eta_{ij}^* = \mathbb{E}(S_{ij} \mid Y_{ijk}; \alpha^*)$

**M step:** ‡ Maximize  $Q(\alpha, \lambda \mid \alpha^*) = l(\eta^*; \alpha) - \frac{1}{2} \alpha^T \mathbf{A}_\lambda \alpha + \frac{1}{2} \log \{|\mathbf{A}_\lambda|_+\}$

- ▶ Estimate  $\alpha$  given the value of  $\lambda$ : P-IRLS

$$\hat{\alpha}_\lambda = \operatorname{argmax}_\alpha \left\{ l(\eta^*; \alpha) - \frac{1}{2} \alpha^T \mathbf{A}_\lambda \alpha \right\}$$

- ▶ Estimate  $\lambda$ : maximize the Laplace-approximated restrictive (or marginal) likelihood

$$L^M(\lambda) = \int \exp \{Q(\alpha, \lambda \mid \alpha^*)\} d\alpha \approx \text{Laplace}(\lambda; \hat{\alpha}_\lambda).$$

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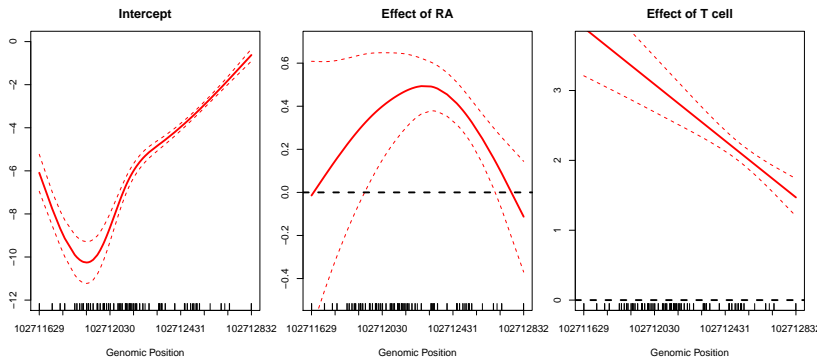
‡ Wood (2011), JRSSB; R package `mgcv`





- ▶ Pointwise confidence intervals
- ▶ Regional tests for non-zero covariate effects
  - ▶ for each covariate, or
  - ▶ for the combined effects of multiple covariates
- ▶ Penalization affects effective degree of freedom

# Results in *BANK1* region



$$p = 1.11e - 16$$

$$p = 6.37e - 218$$

► Error parameters  $p_0 = 0.003$  and  $1 - p_1 = 0.1^{\ddagger}$

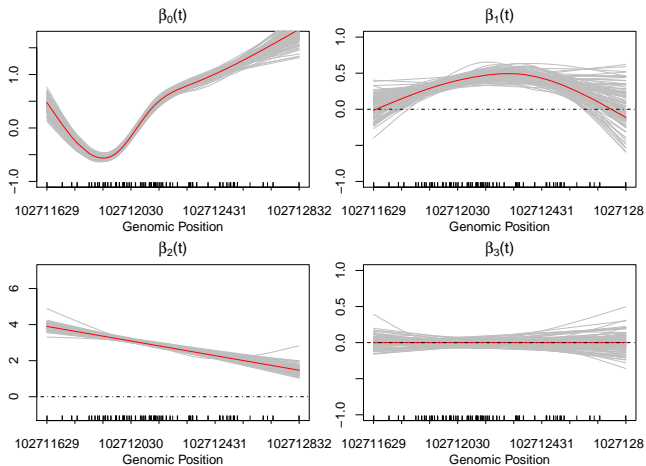
<sup>‡</sup> Prochenka.et al. (2015) *Bioinformatics*.



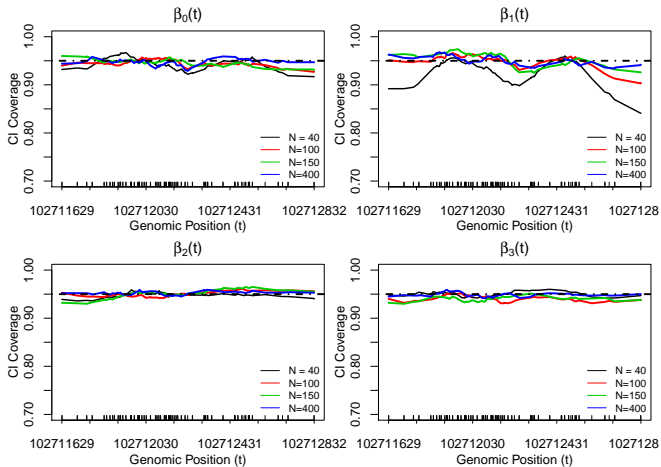
- ▶ Simulated dataset similar to the *BANK1* example
- ▶ One “null” covariate with no effect
- ▶ Two covariates with effects like those seen near *BANK1*
- ▶ Simulate the observed methylated counts  $Y_{ij}$  from

$$Y_{ij} \mid S_{ij} \sim \text{Binomial}(S_{ij}, p_1) + \text{Binomial}(X_{ij} - S_{ij}, p_0).$$

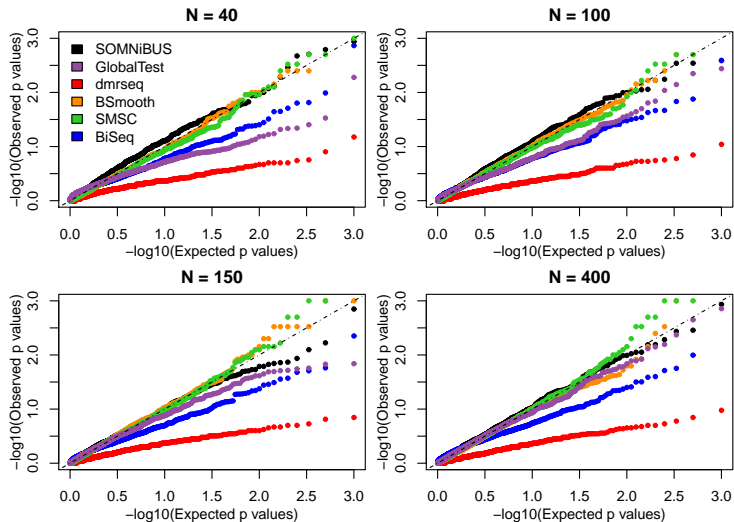
# Little bias in the curve estimates



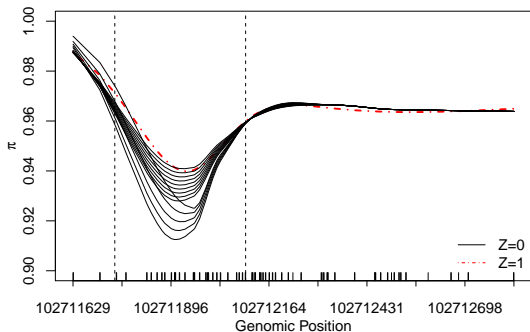
# Empirical confidence interval coverages



# Accurate type I error rates



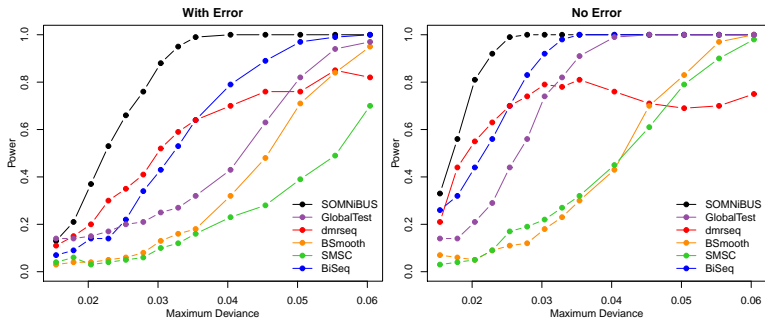
# Simulation to evaluate power



$Z = 1$  curve in red (fixed)

$Z = 0$  curve varied to give various sizes of differences

# Increased power to detect DMRs



Maximum difference between curves

- ▶ With Error:  $p_0 = 0.003, p_1 = 0.9$
- ▶ No Error:  $p_0 = 0, p_1 = 1$





## Advantages

- ▶ Able to use data from many more CpGs where univariate analysis fails / power gain
- ▶ One-stage nature
- ▶ Explicitly allows for experimental errors
- ▶ Inference!



## Advantages

- ▶ Able to use data from many more CpGs where univariate analysis fails / power gain
- ▶ One-stage nature
- ▶ Explicitly allows for experimental errors
- ▶ Inference!

## Room for improvements

- ▶ Its underlying binomial assumption may be overly restrictive
- ▶ It is only applicable for data with negligible (within-group) variability (such as data from inbred animal or cell line experiments)



- ▶ Background and motivation
- ▶ †New method 1: SOMNiBUS (SmOoth ModeliNg of BisUlFite Sequencing)
- ▶ ‡**New method 2: dSOMNiBUS (dispersion-adjusted SmOoth ModeliNg of BisUlFite Sequencing)**

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# Motivating datasets

(from our collaborator Dr. Sasha Bernatsky)



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- ▶ CARTaGENE is an ongoing population-based cohort, including ~43,000 participants aged 40 to 69 years in Quebec
- ▶ The level of anti-citrullinated protein antibodies (ACPA) is a marker of rheumatoid arthritis (RA) risk that often presents prior to any clinical manifestations
- ▶ **Aim:** detect differentially methylated regions (DMRs) associated with ACPA

# Motivating datasets

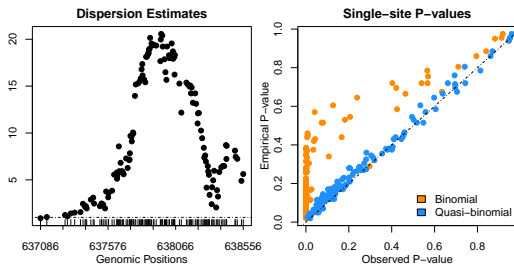
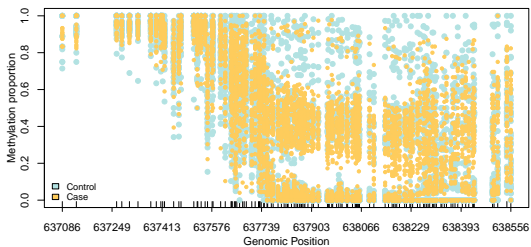
(from our collaborator Dr. Sasha Bernatsky)



- ▶ blood samples of ACPA positive and ACPA negative subjects
  - **covariate of primary interest:** ACPA status
  - **adjusting variables:** age, sex, smoking status and cell type composition(captured by the top 4 PCs)
- ▶ two batches of data, referred to as data 1 and data 2, were collected in 2017 and 2019, respectively.

	data 1 (N =116 )	data 2 (N = 102)
ACPA Positives	55	48
ACPA Negatives	61	54
Number of targeted regions (with at least 50 CpGs)	10,759	12,985

# Observed dispersion in a targeted region



# New method 2: dSOMNiBUS

(dispersion-adjusted SmOoth ModeliNg of Bisulfite Sequencing)



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- ▶ The same error model

$$p_0 = \mathbb{P}(Y_{ijk} = 1 \mid S_{ijk} = 0)$$

$$p_1 = \mathbb{P}(Y_{ijk} = 1 \mid S_{ijk} = 1).$$

- ▶ A quasi-binomial mixed model with the **combination** of
  - a *multiplicative* dispersion,  $\phi$
  - an *additive* dispersion,  $\mathbf{u}$ , (i.e. a subject-specific RE)

$$\log \frac{\pi_{ij}}{1 - \pi_{ij}} = \beta_0(t_{ij}) + \beta_1(t_{ij})Z_{1i} + \beta_2(t_{ij})Z_{2i} + \dots + \beta_P(t_{ij})Z_{Pi} + u_i,$$

$$u_i \stackrel{iid}{\sim} N(0, \sigma_0^2)$$

$$\text{Var}(S_{ij} \mid u_i) = \phi X_{ij} \pi_{ij} (1 - \pi_{ij})$$

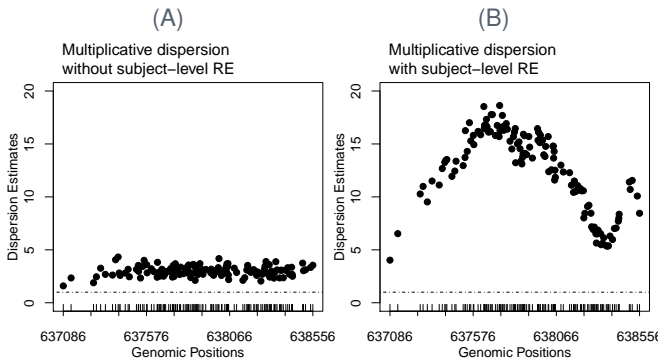
- ▶ Smoothness parameters to penalize the roughness of effect curves.

R package: <https://github.com/kaiqiong/SOMNiBUS>

# RE term enables flexible dispersion patterns in a region



A byproduct of introducing a subject-level RE to a model with smooth covariate effects is a regional dispersion pattern of varying degree.



$$\text{Var}(S_{ij}) \approx X_{ij} \pi_{ij}^* (1 - \pi_{ij}^*) \left\{ \phi + \sigma_0^2 (X_{ij} - \phi) \pi_{ij}^* (1 - \pi_{ij}^*) \right\}$$





- ▶ Three sets of unknown parameters
  - conditional mean parameters (REs):  $\mathcal{B} = (\boldsymbol{\alpha}, \mathbf{u}) \in \mathbb{R}^{N+\sum_0^P L_p}$
  - variance component parameters:  $\Theta = (\boldsymbol{\lambda}, \sigma_0^2) \in \mathbb{R}^{P+2}$
  - **multiplicative dispersion parameter:**  $\phi$
- ▶ The conditional 'distribution' of  $\mathbf{S} \mid \mathcal{B}$  is not available
- ▶ Joint estimation of  $\phi$  and  $\Theta$  is required, as  $\text{Laplace}(\phi, \Theta; \hat{\mathcal{B}}) \neq f(\phi)g(\Theta)$
- ▶ In the presence of data errors, one cannot easily estimate  $\phi$  using the EM algorithm
  - The estimating equation for  $\phi$  is not linear in the unknown methylated counts  $\mathbf{S}$



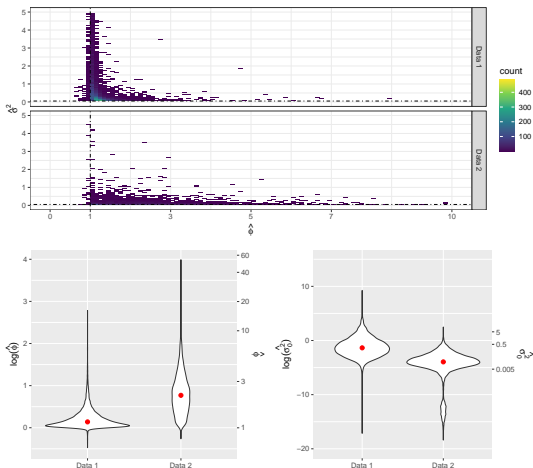
- ▶ Use the notation of **extended quasi-likelihood** to write the conditional quasi-likelihood function
- ▶ Calculate **Laplace-approximated marginal quasi-likelihood** function and its derivatives
- ▶ **A hybrid ES algorithm**
  - A plug-in estimator for  $\phi$  by exploiting its relationship with the dispersion for the contaminated outcome  $\mathbf{Y}$
  - Estimate  $\mathcal{B}$  and  $\Theta$  using ES iterations<sup>‡</sup> assuming  $\phi$  is fixed and known
- ▶ Inference using the observed **quasi-Fisher information**

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<sup>†</sup> Efron (1986), Jorgensen (1987), McCullagh and Nelder (1989)

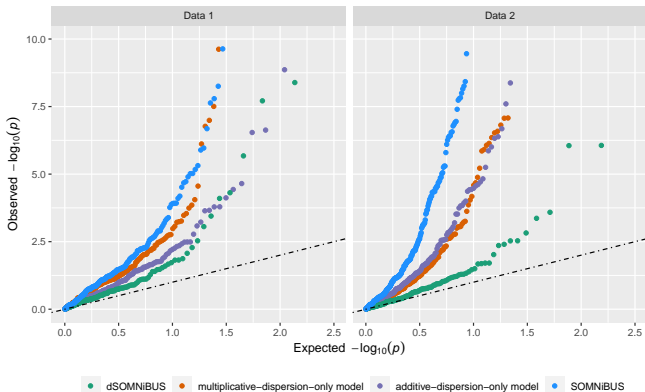
<sup>‡</sup> Elashoff and Ryan (2004)

# Both additive and multiplicative dispersion is present in the data



The distribution of estimated  $\hat{\phi}$  and  $\hat{\sigma}_0^2$  for the 10,759 and 12,985 regions in dataset 1 and 2, respectively.

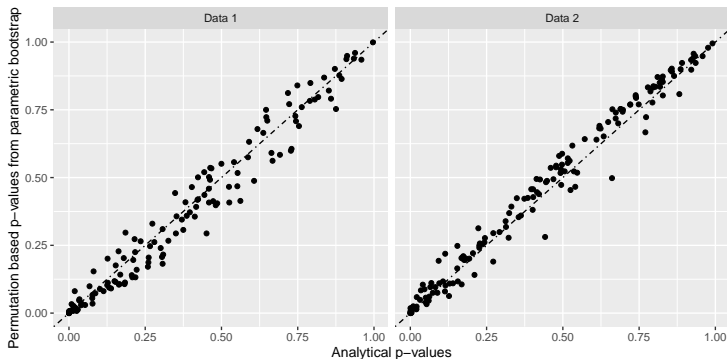
# Ignoring either type of dispersion leads to inflated type I errors



dSOMNiBUS:  $\phi > 0, \sigma_0^2 > 0$ ; multiplicative-dispersion-only model:  $\phi > 0, \sigma_0^2 = 0$

SOMNiBUS:  $\phi = 1, \sigma_0^2 = 0$ ; additive-dispersion-only model:  $\phi = 1, \sigma_0^2 > 0$

# Analytical v.s. bootstrap based p-values



Our inference procedure provides well-calibrated regional p-values.



- ▶ Specify the same  $\beta_p(t)$  and  $Z_p$  as paper 1.

- ▶  $S_{ij} \sim$  **Beta-binomial**  $\left( \mu_{ij} = \pi_{ij}, \rho_{ij} = \frac{\phi - 1}{X_{ij} - 1}, \text{size} = X_{ij} \right)$

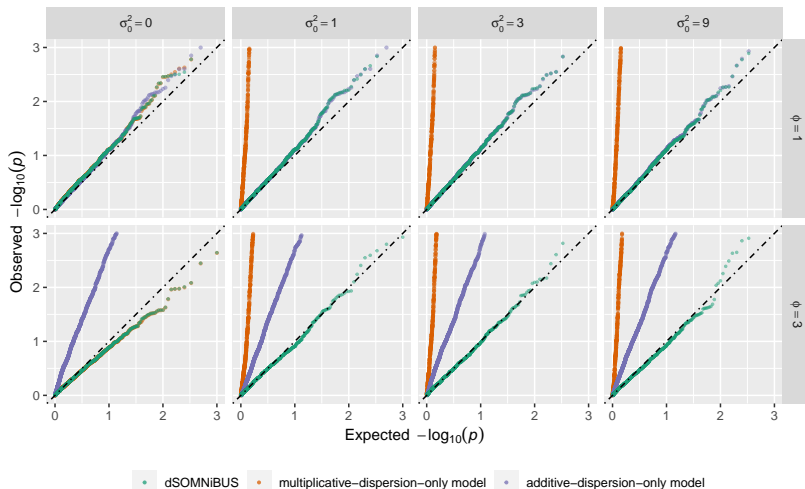
- ▶ In this way, we can always guarantee  $\frac{\text{Var}(S_{ij})}{X_{ij}\pi_{ij}(1 - \pi_{ij})} \equiv \phi$ .

- ▶ Recall: If  $S \sim$  Beta-binomial  $(\mu, \rho, \text{size} = X)$ ,

$$\text{Var}(S) = \underbrace{[1 + (X - 1)\rho]}_{\text{dispersion}} \underbrace{X\mu(1 - \mu)}_{V(\mathbb{E}(Y))}.$$

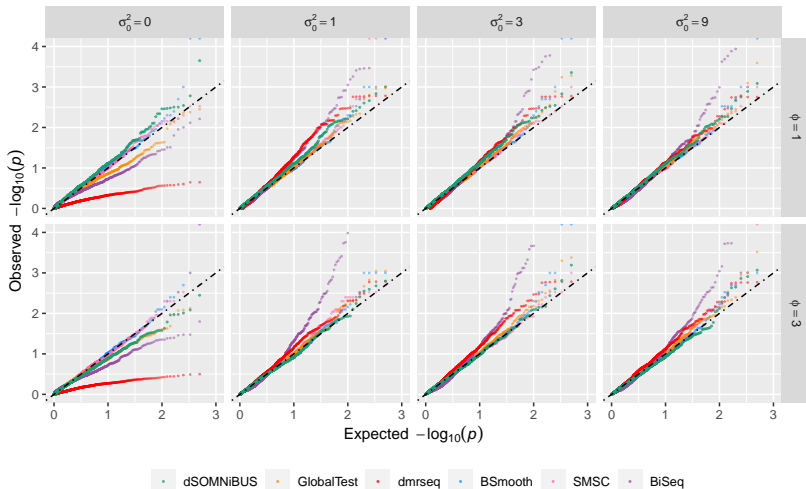
# The impact of dispersion

$$\rho_0 = 0.003, \rho_1 = 0.9$$



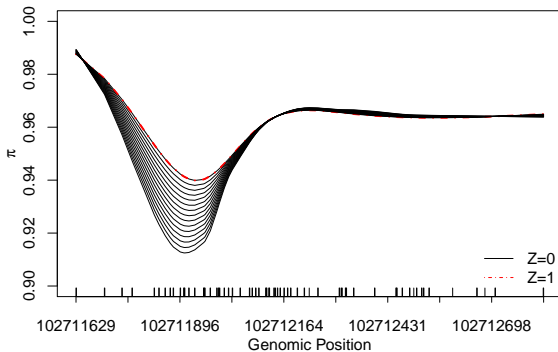
# Type I Error

$\rho_0 = 0.003, \rho_1 = 0.9$





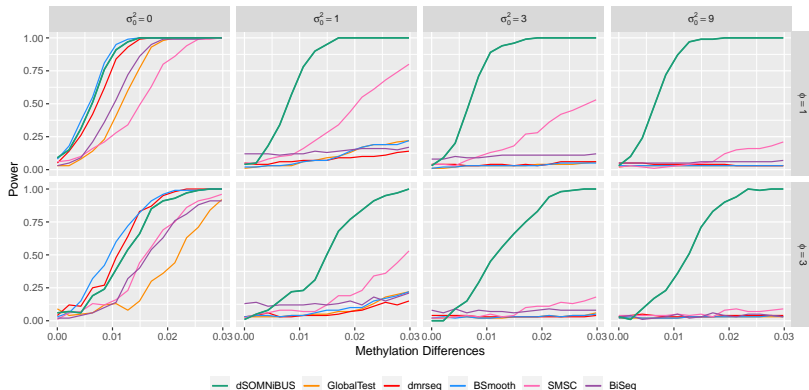
# Simulation to evaluate power



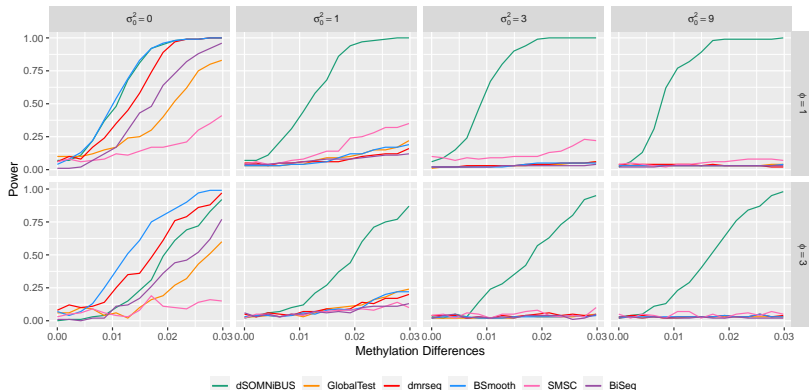
$Z = 1$  curve in red (fixed)

$Z = 0$  curve varied to give various sizes of differences

# Power without errors: $p_0 = 0, p_1 = 1$



# Power with errors: $p_0 = 0.003, p_1 = 0.9$





- ▶ an **adequate representation of realistic dispersion trends** in regional methylation data
- ▶ **well-founded theoretical properties** accounting for all (known) sources of data variability and possible experimental errors
- ▶ **increased power**; correct **control of the type I error rate**
  
- ▶ methodologies can be generally applied to other types of count data
  - allele-specific gene expression (ASE) measured from RNA-seq data
  - any type of count data for a more comprehensive representation of dispersion
  - varying-coefficients models in other context, e.g. temporal trend



- ▶ **integrate SNP information** (automatic variable selection)
- ▶ covariates (eg. disease status) may influence the variability/dispersion of DNA methylation (model  $\phi(Z)$ )
- ▶ correlated samples (additional set of random effects)



## methylation QTL mapping

**Given:** a set of CpGs & a set of nearby SNPs ( $P \gg N$ )

**Output:** genetic variants associated with methylation levels in the test region

### sparseSOMNiBUS

- ▶ a sparsity-smoothness penalty on each functional component  $\beta_p(t)$

$$J(\beta_p) = \lambda \sqrt{(1 - \alpha)J_1(\beta_p) + \alpha J_2(\beta_p)}$$

where

$$J_1(\beta_p) = \int (\beta_p(t))^2 dt$$

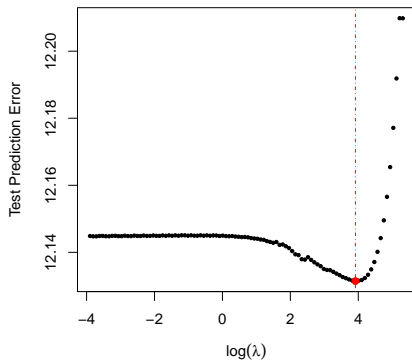
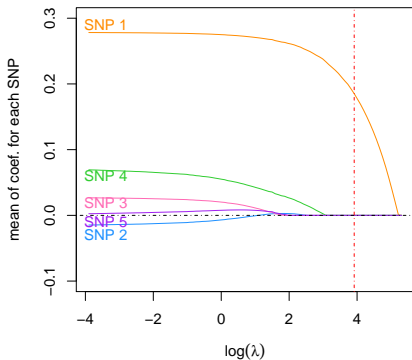
$$J_2(\beta_p) = \int (\beta_p''(t))^2 dt$$

- ▶ proximal gradient descent + backtracking line search (Rcpp)
- ▶ tuning parameters  $\lambda$  and  $\alpha$ , selected by cross-validation

# A simple illustration of sparseSOMNiBUS



- ▶ a methylation region with 123 CpG sites
- ▶ 5 SNPs: 1 mQTL and 4 negative controls



under the best chosen  $\alpha = 0.55$

- ▶ Dr. Celia Greenwood and Dr. Karim Oualkacha
- ▶ Dr. Lajmi Lakhal-Chaieb, Dr. Aurélie Labbe
- ▶ Dr. Yi Yang
- ▶ Dr. Kathleen Klein
- ▶ Dr. Sasha Bernatsky, Dr. Marie Hudson, Dr. Inés Colmegna
- ▶ the CARTaGENE study investigators
- ▶ the participants in the CARTaGENE study







# Thanks

Questions & Comments



- ▶ Assume **known error parameters**  $\rho_0$  and  $\rho_1$ ,

$$\begin{aligned}\rho_0 &= \mathbb{P}(Y_{ijk} = 1 \mid S_{ijk} = 0) \\ \rho_1 &= \mathbb{P}(Y_{ijk} = 1 \mid S_{ijk} = 1).\end{aligned}$$

- ▶ Specify the model

$$\begin{aligned}S_{ij} \mid \mathbf{Z}_i, X_{ij} &\sim \text{Binomial}(X_{ij}, \pi_{ij}) \\ \log \left\{ \frac{\pi_{ij}}{1 - \pi_{ij}} \right\} &= \beta_0(t_{ij}) + \beta_1(t_{ij})\mathbf{Z}_{1i} + \beta_2(t_{ij})\mathbf{Z}_{2i} + \dots + \beta_P(t_{ij})\mathbf{Z}_{Pi},\end{aligned}$$

- ▶ Consider basis expansion:  $\beta_p(t_{ij}) = \sum_{l=1}^{L_p} \alpha_{pl} B_l(t_{ij})$  for  $p = 0, 1, \dots, P$ .
- ▶ <sup>‡</sup>Smoothness parameters to penalize the roughness of effect curves

$$\mathcal{L}^{\text{Smooth}} = \sum_{p=0}^P \lambda_p \int (\beta_p''(t))^2 dt = \sum_{p=0}^P \lambda_p \alpha_p^T \mathbf{A}_p \alpha_p = \alpha^T \mathbf{A} \alpha,$$

<sup>†</sup>R package: <https://github.com/kaiqiong/SOMNiBUS>. <sup>‡</sup>Wahba (1980), Parker and Rice (1985)



## Complete joint likelihood

- ▶ <sup>†</sup> Random-effect view of the smoothness penalty:  $\alpha \sim MVN(\mathbf{0}, \mathbf{A}_\lambda^-)$
- ▶  $l^{\text{complete}}(\mathbf{S}; \alpha, \lambda) = l(\mathbf{S}; \alpha) - \frac{1}{2} \alpha^T \mathbf{A}_\lambda \alpha + \frac{1}{2} \log \{|\mathbf{A}_\lambda|_+\}$

**E step:** Calculate  $\eta_{ij}^* = \mathbb{E}(S_{ij} \mid Y_{ijk}; \alpha^*)$

**M step:** <sup>‡</sup> Maximize  $Q(\alpha, \lambda \mid \alpha^*) = l(\eta^*; \alpha) - \frac{1}{2} \alpha^T \mathbf{A}_\lambda \alpha + \frac{1}{2} \log \{|\mathbf{A}_\lambda|_+\}$

- ▶ Estimate  $\alpha$  given the value of  $\lambda$ : P-IRLS

$$\hat{\alpha}_\lambda = \operatorname{argmax}_\alpha \left\{ l(\eta^*; \alpha) - \frac{1}{2} \alpha^T \mathbf{A}_\lambda \alpha \right\}$$

- ▶ Estimate  $\lambda$ : maximize the Laplace-approximated restrictive likelihood

$$L^M(\lambda) = \int \exp \{Q(\alpha, \lambda \mid \alpha^*)\} d\alpha \approx \text{Laplace}(\lambda; \hat{\alpha}_\lambda).$$

<sup>†</sup> Wahba (1983), JRSSB; Silverman (1985), JRSSB. <sup>‡</sup> Wood (2011), JRSSB; R package `mgcv`



- ▶ Conditional on the values of smoothing parameter  $\lambda$
- ▶ Estimate the variance of EM estimator  $\widehat{\alpha}$ ,  $\mathbf{V}$ , using the observed Fisher information<sup>†</sup>
- ▶ Hypothesis testing for a regional zero effect  $H_0 : \beta_p(t) = 0$ .

- Wald-type statistic

$$T_p = \widehat{\alpha}_p^T \{\mathbf{V}_p\}^{-1} \widehat{\alpha}_p \sim \chi_{\tau_p}^2$$

- Penalization affects effective degree of freedom<sup>‡</sup>;  $\tau_p < L_p = \dim(\alpha_p)$

$$\tau_p = \sum_{l=a_p}^{b_p} (2\mathbf{F} - \mathbf{F}\mathbf{F})_{(l,l)}, \text{ for } p = 0, 1, \dots, P,$$

- $\mathbf{F}$  is the 'hat' matrix and has the form  $\mathbf{F} = (\mathbb{X}^T \widehat{\mathbf{W}} \mathbb{X} + \mathbf{A}_{\widehat{\lambda}})^{-1} \mathbb{X}^T \widehat{\mathbf{W}} \mathbb{X}$

<sup>†</sup> Oakes, D. (1999) Direct calculation of the information matrix via the EM. JRSSB

<sup>‡</sup> Wood, S.N. (2013) On p-values for smooth components of an extended generalized additive model. Biometrika



- ▶ Random-effect view of the smoothness penalty:  $\alpha \sim MVN(\mathbf{0}, \mathbf{A}_\lambda^{-1})$
- ▶ conditional mean parameters (REs):  $\mathcal{B} = (\alpha, \mathbf{u}) \in \mathbb{R}^{N+\Sigma_0^P} L_p$
- ▶ variance component parameters:  $\Theta = (\lambda, \sigma_0^2) \in \mathbb{R}^{P+2}$
- ▶ **multiplicative dispersion parameter:**  $\phi$

## Complete joint log-quasi-likelihood function

$$\begin{aligned}
 q\ell^{(\mathcal{S}, \mathcal{B})}(\mathcal{B}, \phi, \Theta) &= q\ell^{(\mathcal{S}|\mathcal{B})}(\mathcal{B}, \phi) \underbrace{- \frac{1}{2} \alpha^T \mathbf{A}_\lambda \alpha - \frac{1}{2\sigma_0^2} \mathbf{u}^T \mathbf{u}}_{-\frac{1}{2\phi} \mathcal{B}^T \Sigma_\Theta \mathcal{B}} \\
 &\quad + \underbrace{\frac{1}{2} \log \{|\mathbf{A}_\lambda|_+\} + \frac{N}{2} \log (1/\sigma_0^2)}_{1/2 \log \{|\Sigma_\Theta / \phi|_+\}}
 \end{aligned}$$



## Conditional quasi-likelihood function

$$qL^{(S|\mathcal{B})}(\mathcal{B}, \phi) \propto \exp \left\{ -\frac{1}{2\phi} \sum_{i,j} d_{ij}(S_{ij}, \pi_{ij}) - \frac{M}{2} \log \phi \right\},$$

- ▶  $d_{ij}(S_{ij}, \pi_{ij}) = -2 \int_{S_{ij}/X_{ij}}^{\pi_{ij}} \frac{S_{ij} - X_{ij}\pi_{ij}}{\pi_{ij}(1 - \pi_{ij})} d\pi_{ij}$  is the quasi-deviance function
- ▶ This is the extended quasi-likelihood for the joint parameter  $(\mathcal{B}, \phi)$
- ▶ It exhibits the properties of log-likelihood, with respect to both  $\mathcal{B}$  (exact) and  $\phi$  (approximate)
- ▶ †The assumptions required are that  $\phi$  be small and that  $\kappa_r = O(\phi^{r-1})$

† Efron (1986), Jorgensen (1987), McCullagh and Nelder (1989)



## ► Marginal quasi-likelihood function

$$qL^M(\phi, \Theta) = \int \exp \left\{ q\ell^{(\mathbf{S}, \mathbf{B})}(\mathbf{B}, \phi, \Theta) \right\} d\mathbf{B} \approx \text{Laplace}(\phi, \Theta; \hat{\mathbf{B}}) \neq f(\phi)g(\Theta).$$

## ► A similar E-M algorithm

Initialize  $\Theta^{(0)}, \phi^{(0)}, \mathbf{B}^{(0)}$  (estimates ignoring errors); Choose  $\varepsilon = 10^{-6}$ ; Set  $\ell = 0$ ;

**repeat**

- E step:  $\eta_{ij}^{(\ell)} = \mathbb{E}(S_{ij} | Y_{ij}; \mathbf{B}^{(\ell)})$ ;
- M step:  $(\mathbf{B}^{(\ell)}, \phi^{(\ell)}, \Theta^{(\ell)}) = \text{argmax}_{\mathbf{B}, \phi, \Theta} \ell^{\text{Joint}}(\mathbf{B}, \phi, \Theta; \eta_{ij}^{(\ell)})$ . Specifically
  - repeat**
  - Solve  $\mathbf{U}(\mathbf{B}; \Theta^{(s)}) = \mathbf{0}$  to obtain  $\mathbf{B}^{(s)}$  using data  $\eta_{ij}^{(\ell)}$ ;
  - Newton's update for the Laplace approximated marginal likelihood evaluated at data  $\eta_{ij}^{(\ell)}$ :
 
$$(\phi, \Theta)^{(s+1)} = (\phi, \Theta)^{(s)} - \left[ \nabla^2 \text{Laplace}(\mathbf{B}^{(s)}) \right]^{-1} \nabla \text{Laplace}(\mathbf{B}^{(s)});$$

$s \leftarrow s + 1$ ;

**until**  $\|\mathbf{B}^{(s)} - \mathbf{B}^{(s-1)}\|_2 < \varepsilon$ ;

$\ell \leftarrow \ell + 1$ ;

**until**  $\|\mathbf{B}^{(\ell)} - \mathbf{B}^{(\ell-1)}\|_2 < \varepsilon$ ;

Return  $\Theta^{(\ell)}, \mathbf{B}^{(\ell)}, \phi^{(\ell)}$ ;

## ► Estimating $\phi$

- Likelihood-based estimator
- Moment-based estimator (better)



- ▶ Estimate the variance of EM estimator  $\hat{\alpha}$ ,  $\mathbf{V}$ , using the observed (quasi-)Fisher information<sup>†</sup>
- ▶ Hypothesis testing for a regional zero effect  $H_0 : \beta_p(t) = 0$ .
  - Regional statistic

$$T_p = \frac{\hat{\alpha}_p^T \{ \hat{\mathbf{V}}_p \}^{-1} \hat{\alpha}_p}{\tau_p} \sim F_{\tau_p, M-\tau}$$

- $\tau_p$  : EDF for smooth term  $\beta_p(t)$ .  $\tau$  : total EDF of the model
- This F null distribution relies on the assumption that  $(M - \tau)\hat{\phi}/\phi \sim \chi_{M-\tau}^2$ , which is approximately true for moment-based dispersion estimator

<sup>†</sup> Elashoff and Ryan (2004) An EM algorithm for estimating equations. Journal of Computational and Graphical Statistics